

Role of Non-Cultured Keratinocyte Cell Grafting in Take of Full Thickness Skin Graft in Post Burn Contracture

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Abstract

All deeper burns i.e. second degree deep dermal and full thickness heals by scarring that causes restrictions in the movements and aesthetics issues for patients. Burn reconstructive surgery requires that the defects after release should be replaced with donor tissues which have matching texture and colour like autologous skin grafting or flap surgeries. Here we are using this method to look for role in take of FTSG post burn contracture. Full-thickness skin grafts include full thickness of the epidermis and dermis whereas split-thickness skin grafts (STSG) include the entire epidermis and only partial dermis. The main complication of this procedure is risk of graft failure. Keratinocyte cells suspension is claimed to hasten the wound healing. In this article, we share our experience of using non-cultured keratinocyte grafting (NCKG) in improving the take of full thickness skin graft (FTSG).

Keywords: NCKG; FTSG; Post burn contracture.

Introduction

Burn trauma constitutes the second most common cause of trauma related deaths after vehicular accidents, in both developing and developed country. An extensive burn is the most devastating injury that human being had to suffer. After immediate concern for survival in victim, restoration to pre-injury status, and return to daily activities becomes important for victim and treating team.¹ A healed burn patient may be left with contractures and scars with varying degrees of functional issues and cause social stigma among

victims. The healed contractures need to be revised and raw areas covered with FTSG or STSG which may be associated with delayed wound healing of the skin grafts due to aberrant vascularity of the graft bed.

Materials and Methods

This study was conducted in Plastic surgery department in a tertiary care center in the month of November-December 2021. The patient is male child a case of Post burns recurrent band like

constriction of Right index and middle fingers with restriction of daily activities with USS:12/13 (figure 1). Release of post burns contracture with FTSG with K-wire fixation was done. (Figure 2) post operatively there was graft loss (figure 3) for which we have used non cultured keratinocyte epidermal graft (NCKEG) to manage the graft loss. Under all aseptic precautions, a 3cm x 1cm area of groin region was marked (Figure 4) Local anaesthesia (2% xylocaine) was given.

The donor area was derma braded (Figure 4) after the application of mupirocin ointment. The paste, containing dermabraded cells, was collected, homogenized, and was applied on the wound.

A non-adherent dressing was placed on it followed by gauze dressing. The wound was inspected on the 7th day and thereafter weekly. Remnant raw area was calculated on each dressing.



Fig. 1: Post burns contracture in right index and middle finger.



Fig. 2: PBC release with FTSG and K WIRE fixation done.



Fig. 3: FTSG raw area.

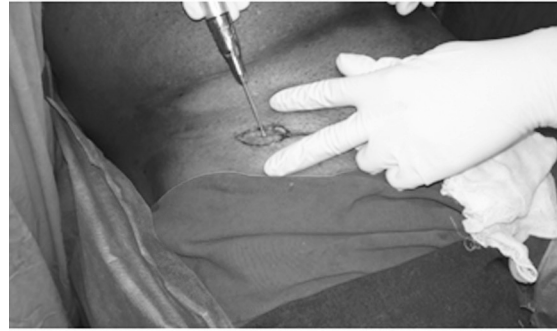


Fig. 4: NCKG harvested.



Fig. 5: Healed FTSG.

Results

NCKG treated wound showed accelerated wound healing (figure 5). Though the grafted cells did not survive but rapid epithelialization started from the periphery of the wound.

Discussion

Wound healing is a complex process. It involves three phases-inflammation proliferation and maturation.¹ The chronic wounds are characterized by a prolonged and persistent proliferative phase due to altered local and systemic factors. The spectrum of modalities available to manage these types of wounds is very wide.

Conveniently it can be grouped into four categories - conventional therapy, novel therapy, reconstructive therapy, and cell-based therapy. Conventional therapies include-conventional dressings with or without topical application of antimicrobial agents, growth factors; various biological dressings such as silver and alginate; hyperbaric oxygen, etc. Novel therapies include the use of platelet-rich plasma, negative pressure wound therapy (NPWT), and skin substitutes. These are minimally invasive with much better

healing efficacy than conventional therapies. Reconstructive therapy, such as skin and flap grafting, are invasive and damage the normal tissue also. Cell-based therapy is also emerging as a part of wound management.^{2,3}

Application of cultured keratinocytes appears to promote healthy granulation tissue formation within the wound bed. The graft, when applied as a sheet, act as an occlusive dressing, preventing wound dehydration and maintaining a moist environment. The majority of evidence suggests that cultured epidermal allografts do not survive indefinitely after transplantation.⁴ Their brief contact with the wound, however, seems sufficient to stimulate reepithelialisation, particularly when dermal tissue is present in the wound bed. This may be due to the release of growth factors by keratinocytes which may favourably influence wound healing. In addition to this, there is a release of several growth factors by keratinocytes that promote wound healing. It is known that cultured keratinocytes release various factors that enhance the growth of other cells in vitro including keratinocytes, fibroblasts, and melanocytes. Identified factors include interleukin-1, other interleukins, and transforming growth factor-alpha.

These keratinocytes may be autologous or allogenic in origin. These cells are separated from skin graft by using trypsin or other methods. After separation, these are cultured in appropriate media to form a sheet. These sheets are used as graft to cover the wound. In our case, we have used autologous non-cultured, non trypsinised keratinocytes cells to promote the healing. We observed favourable

result in terms of formation of healthy granulation tissue and rapid epithelialization of the wound from the margins.

Conclusion

In this study we found that non-cultured keratinocyte grafting has role in healing of the wound and the wound heals at faster rate. But since it is a single case study, definite conclusion cannot be made. Large randomized control trials are required to confirm the efficacy of NCKG in wound healing.

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